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Abstract

Over the past two decades, DNA barcoding has gained popularity in the field of molecular biology as a tool for solving the issue faced by traditional taxonomical methods. Here, we explored the use of this tool to identify various mushroom species found in the diverse ecology of south east Nigeria (Enugu and Ebonyi states). Thus, having an efficient identification system could significantly enhance our ability to make the most use of them for both nutritional and therapeutic values as well as monitor the spatial and temporal patterns of fungal distributions and migrations. By using DNA barcodes from the nuclear ribosomal internal transcribed spacer (ITS) of the rRNA gene, with fungal specific ITS primers (ITS 1 AND 4), ITS barcodes were generated for 71 representative fungal samples, the generated barcodes were sequenced and the result of BLAST identified to 64 samples to species levels with *Lentinus squarrosulus* and *Neonothopanus hygrophanus* as the most abundant species.

Background

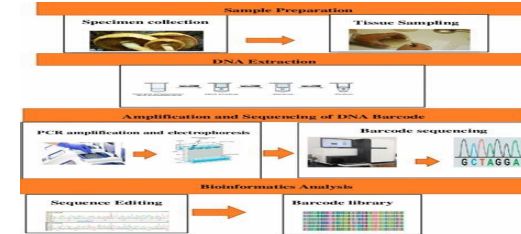
Mushroom is a macro fungus with a distinctive fruiting body, which can be either epigeous or hypogeous and large enough to be seen with naked eye and to be picked by hand. They are widely distributed in the temperate and tropical regions in countries such as Nigeria where they have found importance for use in medicinal and food (Chang Shu-Ting and Philop, 2004). Mushroom are a type of fungi which ubiquitous in both natural and human-made environments. They play important roles in the health of plants, animals, and humans, and in broad ecosystem functions. Adequate knowledge of mushroom diversity and distribution are imperative for successful conservation, management and optimum exploitation of the ecosystem for innumerable benefits to mankind (Nwordu et al. 2013). Mushrooms recognized as visible fungus with characteristic carpophores that symbolize the reproductive stage of several Ascomycetes and Basidiomycetes life cycles. With a variety of nutritional, therapeutic, and ecological benefits, mushrooms constitute a significant bioresource. DNA barcoding is a technique that uses a short DNA sequence to identify species. In the case of fungi, DNA barcoding typically uses the ITS region of the rRNA gene as the barcode region.

Impact and Future Direction

- This investigation serves as a teaching plot and pilot study for undergraduate research where the students gained basic molecular biology skills such as sample collection and documentation, pipetting techniques, gel electrophoresis, DNA extraction and much more.
- There will be a better study involving more students as well as the survey of more sites in the different LGAs in Enugu State to ensure a better representation of the samples size required to publish accurate data on this subject matter.
- Other undergraduate students in the future will use this data as their reference data to compare the nature of species collected and also process for seasonal variations that occur with the change in seasons and time.
- This project will form the background to introduce CURE into the undergraduate curriculum in Nigeria beginning from Godfrey Okoye University, Enugu.

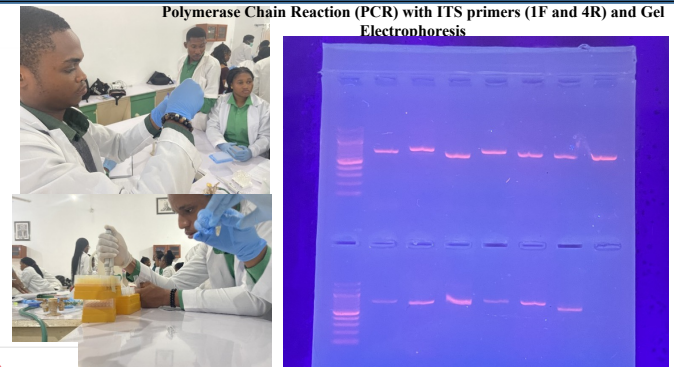
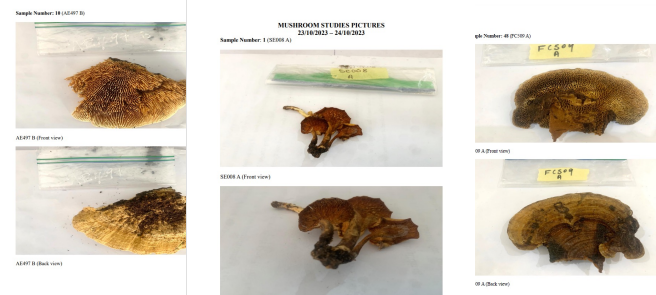
Materials and Methods

The materials used were gloves and ziplock bags during the sample collection. Then we extracted the Mushroom DNA using micropipettes, silica resin, microcentrifuge tubes, centrifuges and a water bath. Then PCR was performed using the thermocycler,. Gel electrophoresis was done then the DNA was sequenced and sent back to us through DNA subway. The last step was bioinformatics.

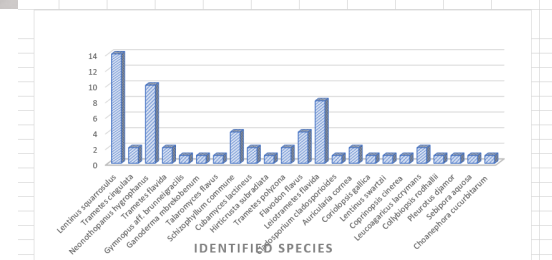
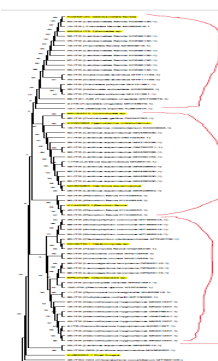


Results

A total of 102 samples were collected



Sample Number	ITS	Species	Accession Number	Count
46	85-ITS	<i>Trametes polyzona</i>	MH131681.1	1
47	87-ITS	<i>Lentinus squarrosulus</i>	OM780271.1	0
48	88-ITS	<i>Lentinus squarrosulus</i>	U9585195.1	0
49	86-ITS	<i>Leucoagaricus leucostriatus</i>	OP985115.1	12
50	90-ITS	<i>Lentinus squarrosulus</i>	MN856300.1	1
51	91-ITS	<i>Lentinus squarrosulus</i>	MN856300.1	1
52	92-ITS	<i>Lentinus squarrosulus</i>	MN856300.1	1
53	93-ITS	<i>Auricularia cornea</i>	MH23359.1	0
54	95-ITS	<i>Leucoagaricus leucostriatus</i>	OP985115.1	12
55	96-ITS	<i>Leucoagaricus leucostriatus</i>	OP985115.1	12
56	97-ITS	<i>Neonothopanus hygrophanus</i>	MK931357.1	1
57	98-ITS	<i>Collybiopsis lodhali</i>	MF100982.1	0
58	99-ITS	<i>Schizophyllum commune</i>	MF406518.1	0
59	100	<i>Pleurotus diamora</i>	K1273359.1	2
60	101 ITS	<i>Sclerophora aquosa</i>	KJ958415.1	2
61	46 ITS	<i>Choanephora cucurbitarum</i>	MF599795.1	1
62	68-ITS	<i>Trametes cingulata</i>	MK736979.1	12
63	94 ITS	<i>Lentinus squarrosulus</i>	MN856300.1	0
64	102 ITS	<i>Neonothopanus hygrophanus</i>	MK931357.1	28



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There are four major Clades - A: comprising majority of genus *Leiotrametes*, Clade B: *Lentinus*, Clade C: *Neonothopanus* and Clad D: *Choanephora* and *Rust Fungus*

CONCLUSIONS

Out of 71 samples, 64 were identified to specie level. These species are in 4 major Clades: Clade A: *Leiotrametes* Complex, Clade B: *Lentinus* Complexes, Clade C: *Neonothopanus* Complexes, Clade D: *Choanephora* and *rust fungus*. The most abundant species were *Lentinus Squarrosulus* and *Neonothopanus hygrophanus*. *Leucoagaricus lacrymans* (samples 89, 95 and 96), *Lentinus squarrosulus* (Sample 33), *Coriolopsis galica* (Sample 65) and *Neonothopanus hygrophanus* (Sample 102) ,had significant mismatches. With reference data on the NCBI reference database. This result conflicts with related research conducted in Cross River and Southwest states of Nigeria , where the main genera found were *Agaricus* and *Aleuria*. on Hence, further bioinformatics studies should be done to understand the nature of these dissimilarities.

Acknowledgements and References

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